



Uni-Fixä

Intended Use

Uni-Fix™ is a general purpose fixative designed to replace formalin as a routine fixative. It is a substitute for 10% Neutral Buffered Formalin.

General Information

Uni-Fix™ is a general purpose fixative designed to replace formalin as a routine fixative. Uni-Fix™ is buffered to produce optimum staining with hematoxylin and eosin. This fixative produces histologic results similar to or better than those following fixation with 10% neutral buffered formalin. Subcellular organelles are well fixed and remain intact, and erythrocytes are not lysed. The fixation artifacts are similar to those obtained with formalin, and staining times for hematoxylin and eosin are the same as that used following fixation with 10% neutral buffered formalin. Uni-Fix™ does not produce the "alcohol artifact" often seen with formalin substitute fixations. This is the routine fixative that should be used in the laboratory to replace 10% neutral buffered formalin. Immunohistochemistry results are similar to those following fixation with formaldehyde.

Packaging

Catalog #	Volume
1400	2oz x 180/cs
1401	15mL x 196/cs
1402	1oz x 144/cs
1403	5mL x 100/cs
1404	4oz x 100/cs
1405	8oz x 48/cs
1409	1 qt
1410	4x1 gal
1420	5 gal ropak

Fixation Procedure

Uni-Fix™ is a coagulative and noncoagulative fixative. It fixes with excellence nuclear and cytoplasmic components of cells. The following information is relevant to the use of Uni-Fix™ as a fixative:

1. Biopsies and fresh tissues should be added directly to Uni-Fix™.
2. No dilution or addition of other agents is necessary before use. Generally it should be used in the same way one uses formalin as a fixative.
3. All biopsies or tissue should be fixed in Uni-Fix™ for at least one hour before processing.
4. Over fixation is not a problem. Large tissues require about the same time for fixation as that used with formalin fixation.
5. No washing of the tissue after fixation is necessary.
6. The fixed tissue should be processed by the same schedule routinely used for formalin-fixed tissues.

7. The schedule for staining tissues fixed with Uni-Fix™ is the same as that for formalin-fixed tissues. The staining times for hematoxylin and eosin will be similar to that used routinely for formalin-fixed tissues. (See attached schedule). The staining times for all special stains are the same as those used routinely for formalin-fixed tissues.
8. Uni-Fix™ contains aldehydes; these should be disposed of similar to the procedure used for formaldehyde. All disposal questions should be directed to your local or county wastewater regulatory agency. Uni-Fix™ contains no mercury or heavy metals.

Staining Procedure

BBC RECOMMENDED AUTOMATED AND MANUAL HISTOLOGY STAINING PROCEDURE FOR HARRIS HEMATOXYLIN AND EOSIN

*Initially deparaffinize tissue sections with BBC S1™ or Xylene

Step *	Solution	Time
1.	100% Alcohol.....	20 seconds
2.	100% Alcohol.....	20 seconds
3.	95% Alcohol.....	20 seconds
4.	95% Alcohol.....	20 seconds
5.	70% Alcohol.....	20 seconds
6.	Running H ₂ O Wash.....	30 seconds
7.	BBC Harris Hematoxylin	4-5 minutes
8.	Running H ₂ O Wash.....	1 minute
9.	BBC Acid Wash•Histo™	1 minute
	or BBC Acid Alcohol•Histo™	2-3 seconds
10.	Running H ₂ O Wash.....	1 minute
11.	BBC Blueing Solution•Histo™.....	15 seconds
12.	Running H ₂ O Wash	1 minute
13.	70% Alcohol.....	30 seconds
14.	BBC Special Eosin I™ or II™, or Eosin Y, or Eosin Y with Phloxine B.....	1 minute
15.	BBC S2•Histo™	20 seconds
17.	BBC S2•Histo™	20 seconds
18.	BBC S2•Histo™	20 seconds
19.	BBC S2•Histo™	20 seconds
20.	BBC S2•Histo™	20 seconds
21.	BBC S3•Histo™ or Xylene.....	20 seconds
22.	BBC S3•Histo™ or Xylene.....	30 seconds
23.	BBC S3•Histo™ or Xylene.....	30 seconds
24.	Mount and coverslip with Optic Mount I™ or an appropriate mounting medium.	

Note: Each of these reagents can be intermixed and used with other staining sequences and other manufacturer's reagents.