



Scott's Tap Water Substitute for Histology

Catalog #4200

Intended Use

Scott's Tap Water substitute for Histology is a blueing reagent designed for histology. It blues hematoxylin gently and minimizes loss of tissue sections and cells from glass slides.

General Information

Scott's Tap Water Substitute for Histology is prepared to supply a gentle but appropriate blueing solution for histology staining. This formulation provides rapid and precise blueing of nuclear chromatin and nuclear membranes. Decreased loss of adherence of tissue sections from glass slides is experienced with Scott's Tap Water Substitute compared with other blueing reagents available. This blueing formulation of Scott's Tap Water Substitute is highly recommended for your routine staining needs in Histology.

Packaging

Catalog #	Volume
4200	1 gal
4201	4x1 gal

Fixation Procedure

Scott's Tap Water can be used as the blueing reagent in the staining procedures for almost any fixative. Consequently, we will describe fixation with the usual standard fixative used in most laboratories, 10% neutral buffered formalin. 10% Neutral Buffered Formalin is a non-coagulative additive fixative. The buffering capacity of our 10% Neutral Buffered Formalin enhances staining by H&E and immunohistochemistry.

1. The biopsies or tissues should be added directly to the 10% Neutral Buffered Formalin. No other dilution or addition of other agents is necessary before use.
2. Small biopsies, such as bone marrow biopsies, should be fixed at least 3 hours prior to processing. Tissue blocks from large tissues, such as lymph nodes or spleen or breast or colon, are best fixed 10-12 hours, although fixation for 46 hours is often sufficient. Over-fixation is not a problem; however, tissues should generally not be fixed longer than one to two weeks.
3. No washing of tissues after fixation is necessary.
4. The fixed tissues should be processed by the standard processing schedules that may vary from one hour to 12 hours. Standard recommended BBC tissue processing schedules are available on request.

5. The schedule for staining tissues fixed with 10% Neutral Buffered Formalin is the similar to the schedule published in standard texts of histology. Our suggested schedule follows.

Staining Procedure

BBC RECOMMENDED AUTOMATED AND MANUAL HISTOLOGY STAINING PROCEDURE FOR HARRIS HEMATOXYLIN AND EOSIN

*Initially deparaffinize tissue sections with BBC S1™ or Xylene

Step *	Solution	Time
1.	100% Alcohol.....	20 seconds
2.	100% Alcohol.....	20 seconds
3.	95% Alcohol.....	20 seconds
4.	95% Alcohol.....	20 seconds
5.	70% Alcohol.....	20 seconds
6.	Running H ₂ O Wash.....	30 seconds
7.	BBC Harris Hematoxylin	3-5 minutes
8.	Running H ₂ O Wash	1 minute
9.	BBC Acid Wash•Histo™	1 minute
	or BBC Acid Alcohol•Histo™	2-3 dips
10.	Running H ₂ O Wash.....	1 minute
11.	BBC Blueing Solution•Histo™	15 seconds
12.	Running H ₂ O Wash.....	1 minute
13.	70% Alcohol.....	30 seconds
14.	BBC Special Eosin I™ or II™, or Eosin Y, or Eosin Y w/ Phloxine B.....	45 seconds
15.	BBC S2•Histo™	20 seconds
16.	BBC S2•Histo™	20 seconds
17.	BBC S2•Histo™	20 seconds
18.	BBC S2•Histo™	20 seconds
19.	BBC S2•Histo™	20 seconds
20.	BBC S3™ or Xylene.....	20 seconds
21.	BBC S3™ or Xylene.....	30 seconds
22.	BBC S3™ or Xylene.....	30 seconds
23.	Mount and coverslip with Optic Mount I™ or an appropriate mounting medium.	

Note: Each of these reagents can be intermixed and used with other staining sequences and other manufacturer's reagents.