



Grossing Aid™

Intended Use

Grossing Aid™ is a fixative designed to enhance the observation of lymph nodes grossly at the dissection or cutting station.

General Information

Grossing Aid™ is an alcoholic formalin with enhancing additives to render tissues with high DNA content white or a lighter color than the surrounding tissues. Grossing Aid™ rapidly fixes adipose tissue and connective tissues. These connective tissues may contain lymph nodes, which in the unfixed state may be difficult to identify. Grossing Aid™ causes lymph nodes to appear white against a darker background of connective tissues and adipose tissues.

Packaging

Catalog #	Volume
0278	1 qt
0280	1 gal
0285	5 gal cube

Fixation Procedure

Grossing Aid™ is a coagulative and non-coagulative additive formaldehyde fixative. It fixes rapidly and can be used as a primary or secondary fixative. Grossing Aid™ is designed specifically to fix hard to fix tissues, such as breast and colon, and tissues that contain large amounts of fat. It is also designed to cause lymph nodes to appear lighter in color grossly than the surrounding tissues.

1. Large specimens received from surgery can be placed in Grossing Aid™ for rapid initial fixation. Tissues that contain large amounts of DNA will turn white. Speed of fixation improves if large specimens are opened prior to fixation. Bowel can be opened; large connective tissue can be crudely serially sectioned.
2. Tissues cut into cassette-sized blocks and enclosed in cassettes can be placed in Grossing Aid™ or in any primary fixative such as 10% NBF, BBC Rapid Fix™ or Stat Fix™ at the cutting station.
3. Tissues primarily fixed in Grossing Aid™ should be processed by the same routine schedule used for formalin-fixed tissues.
4. When used as a primary fixative, over fixation is generally not a problem; however, tissues should not be left in Grossing Aid™ more than one week.
5. Grossing Aid™ is compatible with all other fixatives, and the buffers will not precipitate in subsequent alcohols on the processor.
6. The schedule for staining sections from tissues fixed with Grossing Aid™ is similar to that of formalin-fixed tissues. See the following staining schedule for H&E.

7. Disposal of Grossing Aid™ should be the same as that used for formaldehyde in your laboratory. Grossing Aid™ contains no mercury.

Staining Procedure

BBC RECOMMENDED AUTOMATED AND MANUAL HISTOLOGY STAINING PROCEDURE FOR HARRIS HEMATOXYLIN AND EOSIN

*Initially deparaffinize tissue sections with BBC S3•Histo™ or Xylene

Solution	Time
1. 100% Alcohol.....	20 seconds
2. 100% Alcohol.....	20 seconds
3. 95% Alcohol.....	20 seconds
4. 95% Alcohol.....	20 seconds
5. 70% Alcohol.....	20 seconds
6. Running H ₂ O Wash	30 seconds
7. BBC Harris Hematoxylin	4-5 minutes
8. Running H ₂ O Wash	1 minute
9. BBC Acid Wash•Histo™	1 minute
or BBC Acid Alcohol•Histo™	2-3 seconds
10. Running H ₂ O Wash	1 minute
11. BBC Blueing Solution•Histo™	15 seconds
12. Running H ₂ O Wash	1 minute
13. 70% Alcohol.....	30 seconds
14. BBC Special Eosin I™ or II™, or Eosin Y, or Eosin Y with Phloxine B	1 minute
15. BBC S2•Histo™	20 seconds
17. BBC S2•Histo™	20 seconds
18. BBC S2•Histo™	20 seconds
19. BBC S2•Histo™	20 seconds
20. BBC S2•Histo™	20 seconds
21. BBC S3•Histo™ or Xylene.....	20 seconds
22. BBC S3•Histo™ or Xylene.....	30 seconds
23. BBC S3•Histo™ or Xylene.....	30 seconds
24. Mount and coverslip with Optic Mount I™ or an appropriate mounting medium.	

Note: Each of these reagents can be intermixed and used with other staining sequences and other manufacturer's reagents.