



Tissue Giemsa Stain Kit™

Intended Use

Tissue Giemsa Stain Kit™ is designed to stain with excellence all tissues including fixed bone marrow aspirates, decalcified bone core biopsies and lymph nodes. The colors attained mimic those seen with Romanowsky-type stains of air dried smears.

General Information

Tissue Giemsa Stain Kit™ is designed to product with excellence the standard Romanowsky colors on tissue sections from all tissues, including bone marrow aspirates and lymph nodes. Nuclear chromatin and heterochromatin show distinct staining. Nucleoli are usually dark blue and very visible. Primitive precursor cells of granulocytic and erythrocytic series demonstrate precisely stained chromatin. Cytoplasmic granules, including azurephilic granules of the granulocytic series, are brilliantly stained and vary from purple to red. The cytoplasm of primitive precursor cells is deeply basophilic. Cytoplasmic granules of basophils and mast cells are deep purple. This is the tissue Giemsa stain designed for all hematopathologists and pathologists for enhanced studies of hematopathologic disorders.

Packaging

Catalog #	Volume	
5540	Kit - 1 ea. Solutions A-MM	
5541	Solution A	1 quart
5542	Solution B1	125 mL
5543	Solution B2	1 quart
5544	Solution C	1 quart
5545	Solution MM	125 mL

Fixation Procedure

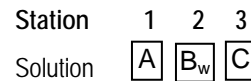
All fixatives for tissues work well. These include 10% NBF, B-Plus Fix™, B5. The decalcifier that works best is Rapid Cal Immuno.

Staining Procedure

Sections of Fixed Bone Marrow Aspirates, Lymph Nodes and Other Tissues

- I. **Prepare Working Solution B(B_w):**
In a coplin jar, add 5 mL of Solution B1 to 45mL of Solution B2
- II. Place 50 mL of each solution in sequentially placed Stations of Coplin jars.
Label them 1, 2, and 3, respectively.

You should now have a staining set up that looks like the following:



- Working solution B_w is good for 5-7 slides or for 1 day.

III. Staining of Tissues:

Deparaffinize and run tissue sections down to tap H₂O

1. **Station 1:** Solution A5 minutes
2. Running Tap H₂O1 minute
3. **Station 2:** Solution B_w
Non decalcified tissues15 minutes
Decalcified tissues20 minutes
4. Running Tap H₂O30 seconds
5. **Station 3:** Solution C.....
Non decalcified tissues15 dips
Decalcified tissues6 dips
6. Running Tap H₂O15 seconds
7. Blot 3 times to remove excess water with lint-less tissue paper (such as Kim Wipes).
8. Drop on Solution MM (Mounting Medium) to cover tissue sections.
9. Blot 5 times to remove excess mounting medium.
10. Let air dry 45 seconds.
11. Place in Xylene or Xylene substitute clearant.
12. Coverslip with Optic Mount I™ or similar Toluene or Xylene based mounting medium. Tape coverslipppers can also be used.