



G.I. Fix™

Intended Use

G.I. Fix™ is formulated specifically for tissues with large amounts of columnar epithelium, such as those from the gastrointestinal tract. G.I. Fix™ is also excellent for needle biopsies of liver.

General Information

G.I. Fix™ is designed specifically for tissues with large amounts of columnar epithelium. It is also ideal for fixing liver biopsies. Following fixation, epithelial cell nuclei show distinct nuclear membranes, precise heterochromatin and euchromatin, and well-defined nucleoli. Neoplastic and reactive cytoplasmic changes in columnar cells demonstrate varying degrees of eosinophilia or basophilia, and goblet cells are well delineated. Cytoplasmic organelles, such as those in Paneth cells and endocrine cells, are distinct and stain precisely. With H&E, cellular membranes stain distinctly and are brightly eosinophilic. This is the ideal fixative for routine fixation of all tissues containing columnar epithelium.

Packaging

Catalog #	Volume
1104	1/2oz, 196/cs
1102	1oz, 144/cs
1100	2oz, 100/cs
1112	1 Quart
1115	4x1 gal
1120	5 Gallon Cube

Fixation Procedure

G.I. Fix™ is a coagulative and noncoagulative fixative. It fixes extensively the nuclear and cytoplasmic components of the mucosa, submucosa, and muscularis of the gastrointestinal tract. It also fixes with excellence hepatic cells, including portal biliary epithelium and inflammatory cells. The following information is relevant to the use of G.I. Fix™:

1. Specimen biopsies, either gastrointestinal or liver, should be promptly placed in G.I. Fix™. No dilution or addition of other agents is necessary before use.
2. G.I. Fix™ should generally be used in the same way one uses 10% neutral buffered Formalin as a fixative.
3. Gastrointestinal biopsies should be fixed in G.I. Fix™ for at least one hour before processing.
4. Over-fixation is not a problem; however, it is a good idea not to fix tissues in G.I. Fix™ for more than one week.

5. Following fixation, washing of tissue is not necessary. Fixed tissue should be processed by the same routine schedule used for formalin-fixed tissues.
6. The schedule for staining sections from tissues fixed with G.I. Fix™ is the same as that of formalin-fixed tissues. See the following staining schedule.
7. Disposal of G.I. Fix™ should be the same as that used for formaldehyde in your laboratory. G.I. Fix™ contains no mercury.

Staining Procedure

BBC RECOMMENDED AUTOMATED AND MANUAL HISTOLOGY STAINING PROCEDURE FOR HARRIS HEMATOXYLIN AND EOSIN

*Initially deparaffinize tissue sections with BBC S3•Histo™ or Xylene

Solution	Time
1. 100% Alcohol.....	20 seconds
2. 100% Alcohol.....	20 seconds
3. 95% Alcohol.....	20 seconds
4. 95% Alcohol.....	20 seconds
5. 70% Alcohol.....	20 seconds
6. Running H ₂ O Wash	30 seconds
7. BBC Harris Hematoxylin.....	4-5 minutes
8. Running H ₂ O Wash	1 minute
9. BBC Acid Wash•Histo™	1 minute
or BBC Acid Alcohol•Histo™	2-3 seconds
10. Running H ₂ O Wash	1 minute
11. BBC Blueing Solution•Histo™	15 seconds
12. Running H ₂ O Wash	1 minute
13. 70% Alcohol.....	30 seconds
14. BBC Special Eosin I™ or II™, or Eosin Y, or Eosin Y with Phloxine B	1 minute
15. BBC S2•Histo™	20 seconds
17. BBC S2•Histo™	20 seconds
18. BBC S2•Histo™	20 seconds
19. BBC S2•Histo™	20 seconds
20. BBC S2•Histo™	20 seconds
21. BBC S3•Histo™ or Xylene.....	20 seconds
22. BBC S3•Histo™ or Xylene.....	30 seconds
23. BBC S3•Histo™ or Xylene.....	30 seconds
24. Mount and coverslip with Optic Mount I™ or an appropriate mounting medium.	

Note: Each or these reagents can be intermixed and used with other staining sequences and other manufacturer's reagents.