# Cyto•Cell Block Fix™

## Intended Use

Cyto•Cell Block Fix™ is designed to fix cells from body fluids for paraffin embedding.

## General Information

Cyto•Cell Block Fix™ is designed to fix with excellence cells from body fluids. Cyto•Cell Block Fix™ causes precipitation of protein and usually causes trapping of cells within the protein matrix present. Alternatively, exogenenous protein, such as Cyto•Clot™ can be added to enhance trapping of cellular elements present. Cyto•Cell Block Fix™ partially erythrocytes to improve paraffin penetration and staining. Nuclei show crisp nuclear membranes with distinct nucleoli. This is the fixative for body fluids that should be used for ideal preparation of paraffin embedded cell blocks.

## Packaging

| Catalog # | Volume |
|---|---|
| 2295 | 5 mL x 100/cs |
| 2300 | 2oz x 100/cs |
| 2305 | 1 qt |
| 2310 | 1 gal |

## Fixation Procedure

1. Dampen the square of a separate piece of lens paper with Cyto•Cell Block Fix™.
2. Inspect the fluid specimen and put any clotted material or pieces of tissue on the separated lens paper and put to one side.
3. Centrifuge approximately 50 cc's of fluid to form a cell button at the bottom of the tube. Pour off the supernatant; then with a pipette re-suspend the cell button.
4. After making smears, add 2-3 cc's of Cyto•Cell Block Fix™ to the sediment remaining in the centrifuge tube.
5. If just a few flakes of sediment are present, add 1 drop of BBC Cyto•Clot™ to the Cyto•Cell Block Fix™. The protein will precipitate out, carrying the flakes with it and adding enough bulk to be visible for embedding. (Cyto•Clot™ can be added to every cell suspension if desired.)
6. Put the funnel in the beaker and line it with the lens paper for filtering.
7. Carefully pipette the sediment into the funnel.
8. Rinse the centrifuge tube thoroughly with Cyto•Cell Block Fix™, washing down the sides, and add to the funnel.
9. When all the Cyto•Cell Block Fix™ has filtered through the funnel, remove the lens paper and spread out the lens paper containing specimen on a Petri dish lid or similar flat object.
10. With the spatula, carefully scrape up the specimen from the funnel filter lens paper and add it to the Cyto•Cell Block Fix™ dampened square of lens paper previously separated.
11. Fold the paper to encase the specimen, enclose it in the cassette, put it in Cyto•Cell Block Fix and give it to the histology department for routine tissue processing.

## Staining Procedure

BBC RECOMMENDED AUTOMATED AND MANUAL HISTOLOGY STAINING PROCEDURE FOR HARRIS HEMATOXYLIN AND EOSIN

*Initially deparaffinize tissue sections with BBC S1™ or Xylene

| Step * | Solution | Time |
|---|---|---|
| 1. | 100% Alcohol | 20 seconds |
| 2. | 100% Alcohol | 20 seconds |
| 3. | 95% Alcohol | 20 seconds |
| 4. | 95% Alcohol | 20 seconds |
| 5. | 70% Alcohol | 20 seconds |
| 6. | Running $H_2O$ Wash | 30 seconds |
| 7. | BBC Harris Hematoxylin | 3-5 minutes |
| 8. | Running $H_2O$ Wash | 1 minute |
| 9. | BBC Acid Wash•Histo™ | 1 minute |
| | **or** BBC Acid Alcohol•Histo™ | 2-3 dips |
| 10. | Running $H_2O$ Wash | 1 minute |
| 11. | BBC Blueing Solution•Histo™ | 15 seconds |
| 12. | Running $H_2O$ Wash | 1 minute |
| 13. | 70% Alcohol | 30 seconds |
| 14. | BBC Special Eosin I™ or II™, **or** Eosin Y, **or** Eosin Y w/ Phloxine B | 45 seconds |
| 15. | BBC S2•Histo™ | 20 seconds |
| 16. | BBC S2•Histo™ | 20 seconds |
| 17. | BBC S2•Histo™ | 20 seconds |
| 18. | BBC S2•Histo™ | 20 seconds |
| 19. | BBC S2•Histo™ | 20 seconds |
| 20. | BBC S3™ or Xylene | 20 seconds |
| 21. | BBC S3™ or Xylene | 30 seconds |
| 22. | BBC S3™ or Xylene | 30 seconds |
| 23. | Mount and coverslip with Optic Mount I™ or an appropriate mounting medium. | |

**Note:** Each or these reagents can be intermixed and used with other staining sequences and other manufacturer's reagents.