



Alcoholic 10% Formalin (3:1 Concentrate) Catalog # 0520

Intended Use

The Alcoholic 10% Formalin (3:1 Concentrate) can be diluted easily to be a working solution of Alcoholic Formalin. Alcoholic 10% Formalin is designed to rapidly fix tissues that are hard to fix, such as fatty tissues, breast and colon. It also functions well as a primary fixative to fix tissues received late in the day and that must be processed quickly overnight. It can also be used on the tissue processor as a secondary fixative in the first station or after the initial fixative station(s).

General Information

Alcoholic 10% Formalin is a rapid fixative designed to be used at the grossing station. It rapidly penetrates tissues prepared at the grossing station and accelerates fixation so that the first fixative on the processor completes the initial fixation process even with large bloody specimens. Alcoholic 10% Formalin is ideal for bowel specimens and breast tissues. Breast, bowel and all fresh tissues benefit from the rapid fixation, which promotes subsequent dehydration and clearing. Additionally, it enhances retention of immunogenicity and improves routine staining with H&E.

Alcoholic 10% Formalin can provide a value added service of rapid pre-fixation prior to definitive cutting of large specimens. Specimens such as bowel and breast, are difficult to cut in the fresh state, and large specimens frequently arrive at the grossing station late in the afternoon. When unfixed, bowel mucosa and submucosa are very soft and moveable and do not attach rigidly to the underlying muscularis propria; consequently, precise cutting of these in cassette-size tissue blocks is difficult. Cutting of tissue blocks can be greatly facilitated by prefixing these specimens with Alcoholic 10% Formalin. Following 1-2 hours in Alcoholic 10% Formalin, bowel and breast can be cut confidently into the proper block sizes. Other large specimens can be prefixed similarly prior to grossing and cutting.

Alcoholic 10% Formalin is compatible with all fixatives in which the tissues have been previously placed. It is also compatible with all dehydrating and clearing reagents. This is the ideal fixative for use at the grossing station for routine fixation, for rapid prefixing of large specimens prior to cutting, and on the tissue processor as a fixative and initial dehydrator.

Packaging

Catalog #	Volume
0520	1 gal
0521	4x1 gal

Fixation Procedure

Alcoholic 10% Formalin is a coagulative and non-coagulative fixative that can be used as a primary or secondary fixative.

As a primary fixative:

- The biopsies or tissues should be added directly to the fixative. Generally it should be used in the same way one uses formalin as a fixative.

- Overfixation is not a problem, and tissues can be left in Alcoholic 10% Formalin for weeks without deleterious effects. It can also be used as a storage medium for tissues following fixation.
- No washing of tissues after fixation is necessary.
- The fixed tissues should be processed by the same schedule used for routine formalin-fixed tissues.
- The staining schedule for tissues fixed with Alcoholic 10% Formalin is the same as that used for formalin fixed tissues.

As a secondary fixative at the cutting (grossing) station:

- Alcoholic 10% Formalin can be used at the grossing station as a secondary fixative. Simply place the prefixed tissues in Alcoholic 10% Formalin following cutting them into cassettes. The tissues can remain in Alcoholic 10% Formalin until placing them on the tissue processor.
- Alcoholic 10% Formalin is compatible with all other fixatives.

As a secondary fixative on the tissue processor:

- Alcoholic 10% Formalin can be used on the tissues processor either in the initial station, or following other fixatives and prior to the dehydrant. Alcoholic 10% Formalin will also begin dehydration gently.
- Alcoholic 10% Formalin is compatible with all other fixatives and with all dehydrants used on the tissue processor.

Staining Procedure

BBC RECOMMENDED AUTOMATED AND MANUAL HISTOLOGY STAINING PROCEDURE FOR HARRIS HEMATOXYLIN AND EOSIN

*Initially deparaffinize tissue sections with BBC S1™ or Xylene

Step	Solution	Time
1.	100% Alcohol	20 seconds
2.	100% Alcohol	20 seconds
3.	95% Alcohol	20 seconds
4.	95% Alcohol	20 seconds
5.	70% Alcohol	20 seconds
6.	Running H ₂ O Wash	30 seconds
7.	BBC Harris Hematoxylin	3-5 minutes
8.	Running H ₂ O Wash	1 minute
9.	BBC Acid Wash-Histo™ or BBC Acid Alcohol-Histo™	1 minute
10.	Running H ₂ O Wash	1 minute
11.	BBC Blueing Solution-Histo™	15 seconds
12.	Running H ₂ O Wash	1 minute
13.	70% Alcohol	30 seconds
14.	BBC Special Eosin I™ or II™, or Eosin Y, or Eosin Y with Phloxine B..	45 seconds
15.	BBC S2•Histo™	20 seconds
17.	BBC S2•Histo™	20 seconds
18.	BBC S2•Histo™	20 seconds
19.	BBC S2•Histo™	20 seconds
20.	BBC S2•Histo™	20 seconds
21.	BBC S3•Histo™ or xylene	20 seconds
22.	BBC S3•Histo™ or xylene	30 seconds
23.	BBC S3•Histo™ or xylene	30 seconds
24.	Mount and coverslip with Optic Mount I™ or an appropriate mounting medium.	

Note: Each of these reagents can be intermixed and used with other staining sequences and other manufacturer's reagents.