

# Acid Alcohol•Histo™

Catalog #3800

## Intended Use

Acid Alcohol•Histo™ is formulated to be used as a differentiator for Hematoxylin in regressive staining procedures. It produces excellent differentiation of nuclei and non-nuclear structures.

## General Information

Acid Alcohol•Histo™ is designed as a differentiator of Hematoxylin in regressive H&E staining procedures. This differentiator provides ideal differentiation of nuclear structures. Non-nuclear structures show clear staining by eosin with precise color variations by the eosin. Residual mucin staining by the hematoxylin is minimal to slight. It is an excellent differentiator of BBC Harris Hematoxylin.

## Packaging

Catalog #	Volume
3810	4x1 gal

## Fixation Procedure

Acid Alcohol•Histo™ is a differentiator of hematoxylin used in H&E staining procedures. It works well with tissues fixed in virtually any type of fixative. The time of differentiation of Acid Alcohol•Histo™ should not vary regardless of the fixative used.

Because Acid Alcohol•Histo™ differentiates hematoxylin precisely regardless of the fixative used, the fixation procedure for 10% neutral buffered formalin will be given.

10% Neutral Buffered Formalin is a non-coagulative additive fixative. The buffering capacity of our 10% Neutral Buffered Formalin enhances staining by H&E and immunohistochemistry.

1. The biopsies or tissues should be added directly to the 10% Neutral Buffered Formalin. No dilution or addition of other agents is necessary before use.
2. Small biopsies, such as bone marrow biopsies, should be fixed at least 3 hours prior to processing.
3. Large tissues, such as tissue blocks from lymph nodes, spleen, breast or colon, are best fixed 10-12 hours, although fixation for 4-6 hours is often sufficient. Over-fixation is not a problem; however, tissues should generally not be fixed longer than one to two weeks.
4. No washing of tissues after fixation is necessary.
5. The fixed tissues should be processed by standard processing schedules that may vary from one to 12 hours. Standard recommended BBC tissue processing schedules are available on request.

6. The schedule for staining tissues fixed with 10% Neutral Buffered Formalin is the same standard schedule published in standard texts of histology. Our suggested schedule follows.
7. Disposal 10% Neutral Buffered Formalin should be the same as that used for fixatives containing formaldehyde. Consult your local wastewater disposal authority for specific instructions.

## Staining Procedure

BBC RECOMMENDED AUTOMATED AND MANUAL HISTOLOGY STAINING PROCEDURE FOR HARRIS HEMATOXYLIN AND EOSIN

\*Initially deparaffinize tissue sections with BBC S1™ or Xylene

Step *	Solution	Time
1.	100% Alcohol.....	20 seconds
2.	100% Alcohol.....	20 seconds
3.	95% Alcohol.....	20 seconds
4.	95% Alcohol.....	20 seconds
5.	70% Alcohol.....	20 seconds
6.	Running H <sub>2</sub> O Wash .....	30 seconds
7.	BBC Harris Hematoxylin .....	3-5 minutes
8.	Running H <sub>2</sub> O Wash .....	1 minute
9.	BBC Acid Wash•Histo™ .....	1 minute
	or BBC Acid Alcohol•Histo™ ....	2-3 dips
10.	Running H <sub>2</sub> O Wash .....	1 minute
11.	BBC Blueing Solution•Histo™ .....	15 seconds
12.	Running H <sub>2</sub> O Wash .....	1 minute
13.	70% Alcohol.....	30 seconds
14.	BBC Special Eosin I™ or II™, or Eosin Y, or Eosin Y w/ Phloxine B.....	45 seconds
15.	BBC S2•Histo™ .....	20 seconds
16.	BBC S2•Histo™ .....	20 seconds
17.	BBC S2•Histo™ .....	20 seconds
18.	BBC S2•Histo™ .....	20 seconds
19.	BBC S2•Histo™ .....	20 seconds
20.	BBC S3™ or Xylene .....	20 seconds
21.	BBC S3™ or Xylene .....	30 seconds
22.	BBC S3™ or Xylene .....	30 seconds
23.	Mount and coverslip with Optic Mount I™ or an appropriate mounting medium.	

**Note:** Each of these reagents can be intermixed and used with other staining sequences and other manufacturer's reagents.