



Zinc Formalin

Intended Use

Zinc Formalin is designed for all tissues. It enhances nuclear and cytoplasmic fixation. BBC Zinc Formalin is buffered ideally to promote excellent H&E staining and to enhance immunohistochemistry results.

General Information

Zinc Formalin is prepared to serve as an excellent fixative for routine light microscopy. It has the non-coagulant fixative qualities of formalin and the coagulant fixative properties provided by zinc. Additionally, it preserves the immunoreactivity of tissue antigens better than neutral buffered formalin alone. This is an excellent fixative for all tissues processed in the histology laboratory. Following fixation, the processing and staining times remain the same as those used after fixation with 10% neutral buffered formalin.

Packaging

Catalog # Volume

0800	2oz, 100/cs
0801	5mL, 100/cs
0802	1/2oz, 100/cs
0803	15mL, 100/cs
0804	1oz, 100/cs
0805	4oz, 100/cs
0810	1 qt
0820	1 gal
0822	5 gal cube

Fixation Procedure

Zinc Formalin is a fixative that contains coagulative and non-coagulative fixatives. It is intended to fix all tissues with excellence.

1. The tissues should be added directly to the Zinc Formalin. No dilution or addition of other agents is necessary before use.
2. Zinc Formalin should be used in the same way one uses formalin as a fixative.
3. Small tissues, such as biopsies from selected organs, should be fixed at least 3 hours prior to processing. Large tissues are best fixed 10-12 hours or overnight. Overfixation is generally not a problem; however, tissues should generally not be fixed longer than one week. Storage of tissues is best done in 70% ethanol.
4. No washing of tissues after fixation is necessary.
5. The fixed tissues should be processed by the same schedule used for routine formalin-fixed tissues.

6. The schedule for H & E staining of sections of tissues fixed with Zinc Formalin is the same as that used routinely for formalin fixed tissues.
7. Disposal of Zinc Formalin should be the same as that utilized for fixative containing formaldehyde.

Staining Procedure

BBC RECOMMENDED AUTOMATED AND MANUAL HISTOLOGY STAINING PROCEDURE FOR HARRIS HEMATOXYLIN AND EOSIN

*Initially deparaffinize tissue sections with BBC S1™ or Xylene

Step *	Solution	Time
1.	100% Alcohol.....	20 seconds
2.	100% Alcohol.....	20 seconds
3.	95% Alcohol	20 seconds
4.	95% Alcohol.....	20 seconds
5.	70% Alcohol	20 seconds
6.	Running H ₂ O Wash	30 seconds
7.	BBC Harris Hematoxylin.....	3-5 minutes
8.	Running H ₂ O Wash	1 minute
9.	BBC Acid Wash-Histo™ or BBC Acid Alcohol-Histo™	1 minute
10.	Running H ₂ O Wash	1 minute
11.	BBC Blueing Solution-Histo™	15 seconds
12.	Running H ₂ O Wash	1 minute
13.	70% Alcohol.....	30 seconds
14.	BBC Special Eosin I™ or II™, or Eosin Y, or Eosin Y with Phloxine B..	45 seconds
15.	BBC S2•Histo™	20 seconds
16.	BBC S2•Histo™	20 seconds
17.	BBC S2•Histo™	20 seconds
18.	BBC S2•Histo™	20 seconds
19.	BBC S2•Histo™	20 seconds
20.	BBC S3•Histo™ or Xylene.....	20 seconds
21.	BBC S3•Histo™ or Xylene.....	30 seconds
22.	BBC S3•Histo™ or Xylene.....	30 seconds
23.	Mount and coverslip with Optic Mount I™ or an appropriate mounting medium.	

Note: Each or these reagents can be intermixed and used with other staining sequences and other manufacturer's reagents.